

# ISOLATION OF SOME MICROALGAE STRAINS BELONGING TO THE FAMILY SCENEDESMACEAS (CHLOROPHYTA) IN FRESHWATER BODIES IN CENTRAL VIETNAM AND INVESTIGATION INTO THEIR BIOLOGICAL CHARACTERISTICS

Tran Nguyen Quynh Anh, Vo Van Minh, Tran Ngoc Son, Trinh Dang Mau\*

The University of Danang, University of Science and Education; tdm@ued.udn.vn

**Abstract** - Seven microalgae strains from samples collected from freshwater lakes in Central Vietnam were isolated and cultivated in BBM medium for investigating their biological characteristics and their applicability. These strains were identified belonging to 6 species, 3 genera of the family Scenedesmaceae, division Chlorophyta. Their average growth rate was from 0.081 - 0.160 d<sup>-1</sup>, and the highest cell density of each strain reached from 2.6 x 10<sup>6</sup> - 60 x 10<sup>6</sup> cells/ml. The total content of pigments (including chlorophyll-a, chlorophyll-b, and carotenoid) of all strains was not too high (4.10 - 11.3 µg/mL), but the total lipid content exhibiting noticeably high levels, accounted for 16.6% - 74.1% of dry cell weight. Among isolated strains, *Desmodesmus abundans* and *D. costato-granulatus* were the most prominent with the highest growth rate and density in combination with remarkably high contents of lipids and pigments, and thus were identified as two potentially great candidates for biofuel production and chlorophyll and carotenoid extraction.

**Key words** - Microalgae; Scenedesmaceae; isolate; biological characteristics; lipid

## 1. Introduction

Scenedesmaceae is a family of green algae, in the class Chlorophyceae, division Chlorophyta. It is one of the most common microalgae families in freshwater bodies that plays many important roles in aquatic ecosystems. Currently, many species of this family have been reported to be effective as a biological source for producing food, nutritional supplements, and drugs for fish and humans since they contain a lot of nutrients including protein, carbohydrates, lipids, vitamins, as well as antioxidant agents, etc. [1], [2]. Sixteen species of genus *Scenedesmus* in Himalaya, India were isolated and exploited in fisheries and a single-cell protein source [3]. Furthermore, some microalgae strains such as strains belonging to the genus *Scenedesmus* have been reported to be effective for the biological treatment of polluted water [4], [5]. Meanwhile, the other *Scenedesmus* species, especially *Scenedesmus obliquus* were found to have the potential to produce high quality biodiesel that meets the modern industry standards [6], [7], [8]. With such important findings, Scenedesmaceae species continue to be an interesting object of many studies in the world with efforts to isolate and find out more strains for practical applications.

In Vietnam, Scenedesmaceae was also a rather common family in freshwater bodies [9], [10], [11], and some species were even recorded to be dominant in river ecosystems [12]. This is a great valuable potential resource for Vietnam but the information about these species has been very limited. This study aims at isolating some microalgae strains of the Scenedesmaceae family from some freshwater bodies in Central Vietnam and investigating their biological characteristics in order to provide scientific data on this group and evaluate their potential for practical application in Vietnam.

## 2. Materials and methods

Microalgae samples were collected by a phytoplankton net with a mesh size of 30 µm in three lakes in Da Nang city (Lake Thac Gian - Vinh Trung, Lake Cong vien 29/3, and Lake Bau Trang) and one lake in Thua Thien Hue province (Lake Bau Thiem). Water environmental characteristics (including pH, DO, TDS, NO<sub>2</sub><sup>-</sup>, NO<sub>3</sub><sup>-</sup>, PO<sub>4</sub><sup>3-</sup>) at the sampling sites were also identified.

Microalgae samples were grown with sterilized bold basal medium (BBM) [13] under a temperature of 25°C, the fluorescent light intensity of 2500 lux, and day/night lighting time of 16:8. The Scenedesmaceae strains were isolated by the method of inoculation of single colonies on agar with BBM in Petri dishes. The complete purified microalgae species were finally separated and continued growing in 5 mL vials containing BBM medium. Isolated microalgae strains were observed and photographed by a Hund (H6000) microscope and identified by morphological comparison. The major works used for the identification were from Bellinger & Sigeo [14] and Tien & Hanh [9].

Spectrophotometric absorption method was used to quantify the density and growth rate of microalgae. Optical density (OD) measurements were carried out at these two wavelengths using a Jasco V-750 UV/VIS Spectrophotometer. Growth rate µ (d<sup>-1</sup>) of microalgae was calculated according to the following equation [15]:

$$\mu = \frac{\ln(Nt) - \ln(N0)}{(T_t - T_0)}$$

Where, Nt and N0: cell density at T<sub>t</sub> and T<sub>0</sub>, respectively (cell/mL); T: time (day).

Generation time was calculated as follows [16]:

$$g = \frac{1}{k} \text{ (hour/generation)}$$

$$\text{with, } k = \frac{\log N_2 - \log N_1}{0.301 h}$$

where, N<sub>2</sub>, N<sub>1</sub>: cell density at T<sub>2</sub> and T<sub>1</sub>(cell/mL);

h: period between T<sub>2</sub> and T<sub>1</sub>(hour).

The extraction procedure of chlorophyll and carotenoid content was conducted with methanol 90%. The mixture of microalgae cells and methanol was incubated in the dark for 24 hours and was centrifuged for 30 min at 4,000 rpm. The obtained supernatant was then measured by spectrophotometer UV-Vis at 470 nm, 652 nm, and 665 nm. Concentrations of chlorophyll-a, chlorophyll-b, and total carotenoid were calculated as follows [17]:

Chlorophyll-a (Ch-a) =  $16.72(A_{665}) - 9.16(A_{652})$

Chlorophyll-b (Ch-b) =  $34.09(A_{652}) - 15.28(A_{665})$

Total carotenoid content

$$(C_{x+c}) = [1000(A_{470}) - 1.63Ch-a - 104.96Ch-b] / 221$$

(Where A: Absorbance)

Lipid extraction and purification were carried out following the method of Bligh and Dyer [18]. Briefly, 15 mL of microalgae cells were mixed with 2 mL of methanol and 1 mL of chloroform before being incubated for 24 hours at 25°C in the dark. The mixture was then mixed with 1 mL of chloroform and 1.8 mL of distilled water and centrifuged at 2,000 rpm for 10 minutes to separate into layers. The lower layer was filtered through a filter paper (Whatman, 0.47 µm pore-size) and the supernatant was weighed (W1). Then, the supernatant was dried at 104°C for 30 minutes to achieve a constant weight (W2). Total lipid content (expressed as a % of dry cell weight (% dcw)) was calculated by subtracting W1 from W2.

Data analysis was performed with R software.

### 3. Results and discussion

#### 3.1. Morphological characteristics of isolated microalgae strains

The isolation results recorded 7 strains belonging to 6 species, 3 genera of the family Scenedesmaceae, division Chlorophyta (Table 1).

Table 1. List of isolated microalgae species composition

<b>Division Chlorophyta</b>
<b>Class Chlorophyceae</b>
<b>Order Sphaeropleales</b>
<b>Family Scenedesmaceae</b>
<b>Genus Scenedesmus</b>
1. <i>Scenedesmus quadricauda</i> C2 (Turpin) Brébisson 1835
2. <i>Scenedesmus quadricauda</i> C3 (Turpin) Brébisson 1835
<b>Genus Desmodesmus</b>
3. <i>Desmodesmus abundans</i> (Kirchner) E.H.Hegewald 2000
4. <i>Desmodesmus bicaudatus</i> (Dedusenko) P.M.Tsarenko 2000
5. <i>Desmodesmus costato-granulatus</i> (Skujala) E.Hegewald 2000
<b>Genus Tetradesmus</b>
6. <i>Tetradesmus obliquus</i> (Turpin) MJWynne 2016
7. <i>Tetradesmus incrassatulus</i> (Bohlin) MJWynne 2016

***Tetradesmus obliquus* (Turpin) MJWynne 2016** (Figure 1-a)

**Description:** Coenobium consists of 2 cells arranging linearly and tying together at the middle of the cells. The cell is rhombic with pointed heads. Cell walls are smooth, with 1 starch granule. The cell size is  $13.66 \pm 1.27$  µm in length and  $5.15 \pm 1.29$  µm in width.

**Ecology and distribution:** Lake Bau Thiem, Thua Thien Hue province. Environmental characteristics at the sampling site are pH = 7.19,  $PO_4^{3-} = 0.340$  mg/L,  $NO_3^- = 0.62$  mg/L,  $NO_2^- = 0.021$  mg/L, TDS = 1.4 mg/L, DO = 6.77 mg/L. This species has been recorded widely distributed in different types of water bodies, especially in

moderate polluted water bodies.

***Desmodesmus abundans* (Kirchner) E. H. Hegewald 2000** (Figure 1-b)

**Description:** This species exists in unicellular form or coenobium with 2 - 4 cylindrically oval cells arranged in a row, outer cells with 1 spine at the outer side and 2 spines at both ends which are  $5.77 \pm 0.42$  µm long (reaching a length up to 15 - 18 µm). Cell size is  $8.31 \pm 0.74$  µm in length and  $3.55 \pm 0.27$  µm in width.

**Ecology and distribution:** Lake Thac Gian - Vinh Trung, Da Nang city. Environmental characteristics at the sampling site: pH = 6.92,  $PO_4^{3-} = 0.131$  mg/L,  $NO_3^- = 0.148$  mg/L,  $NO_2^- = 0.028$  mg/L, TDS = 519 mg/L, DO = 1.85 mg/L. This species has been recorded widely distributed in different water bodies in Europe, North and South America, South Africa and Asia.

***Tetradesmus incrassatulus* (Bohlin) MJWynne 2016** (Figure 1-c)

**Description:** Coenobium of 2 - 4 cells arranging linearly. The cell is rhombic, and round at both ends, with 1 pyrenoids, spines were not observed. The cell size is  $110.69 \pm 0.86$  µm in length and  $5.66 \pm 0.69$  µm in width.

**Ecology and distribution:** Lake Cong Vien 29/3, Da Nang city. Environmental characteristics at the sampling site were aforementioned.

***Scenedesmus quadricauda* C2 (Turpin) Brébisson 1835** (Figure 1-d)

**Description:** Coenobium consists of 2 - 4 - 8 cells arranged in a row. The cells are cylindrical and slightly pointed at both ends. Outer cells have 2 slightly curved inward spines at both ends. The cell wall is smooth, with 1 pyrenoids. The cell length is  $14.85 \pm 0.64$  µm and cell width is  $5.41 \pm 0.22$  µm. The spines length is  $11.27 \pm 0.54$  µm.

**Ecology and distribution:** Lake Thac Gian - Vinh Trung, Da Nang city. Environmental characteristics at the sampling site were already mentioned above. This species has been recorded widely distributed in both floating form and benthic form in all types of water bodies all over the world.

***Desmodesmus bicaudatus* (Dedusenko) P. M. Tsarenko 2000** (Figure 1-e)

**Description:** Coenobium is with 2 linear arranged cells. The cells are nearly ellipsoidal, with 1 straight spine ( $7.22 \pm 0.24$  µm length) at one end of each outer cell, cell walls are smooth. The cell size is  $10.38 \pm 0.07$  µm in length and  $3.66 \pm 0.31$  µm in width.

**Ecology and distribution:** Lake Cong Vien 29/3, Da Nang city. Environmental characteristics at the sampling site are pH = 6.92,  $PO_4^{3-} = 0.085$  mg/L,  $NO_3^- = 0.128$  mg/L,  $NO_2^- = 0.115$  mg/L, TDS = 460 mg/L, DO = 2.59 mg/L. This species has been recorded in several water bodies in Siberia (Russia), Europe (France, Hungary), West Africa, Asia.

***Scenedesmus quadricauda* C3 (Turpin) Brébisson 1835** (Figure 1-f)

**Description:** Coenobium consists of 2 - 4 - 8 ellipsoidal

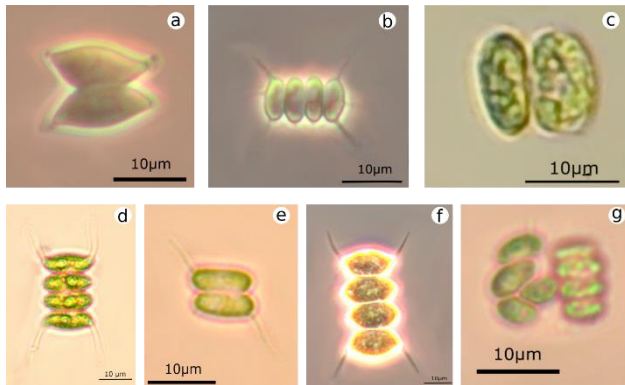
cells arranging linearly. Outer cells have 2 spines at both ends. The cell size is  $19.88 \pm 1.05 \mu\text{m}$  in length and  $10.26 \pm 0.58 \mu\text{m}$  in width. The length of spines is  $11.27 \pm 0.54 \mu\text{m}$ .

**Ecology and distribution:** Lake Thac Gian - Vinh Trung, Da Nang city. Environmental characteristics at the sampling site were aforementioned. This species has been recorded widely distributed in floating form and benthic form of all types of water bodies all over the world.

***Desmodesmus costato-granulatus* (Skuja) E. Hegewald 2000** (Figure 1-g)

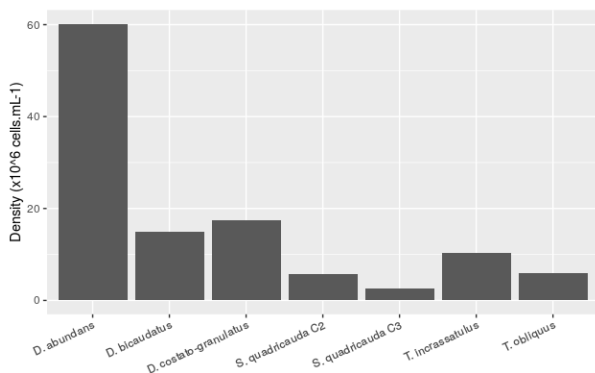
**Description:** Coenobium may form or not, consists of 1 - 2 - 4 cells arranging linearly. The cells are egg shaped or elongated egg shape; spines were not observed. Dimensions of cells are  $5.54 \pm 0.46 \mu\text{m}$  length and  $1.96 \pm 0.24 \mu\text{m}$  width.

**Ecology and distribution:** Lake Bau Trang, Da Nang city. Environmental characteristics at the sampling site are  $\text{pH} = 6.93$ ,  $\text{PO}_4^{3-} = 0.078 \text{ mg/L}$ ,  $\text{NO}_3^- = 0.31 \text{ mg/L}$ ,  $\text{NO}_2^- = 0.028 \text{ mg/L}$ ,  $\text{TDS} = 7809 \text{ mg/L}$ ,  $\text{DO} = 6 \text{ mg/L}$ . This species has been recorded to be distributed in a wide range of nutritional status of water bodies, from oligotrophic to eutrophic water bodies.



**Figure 1.** Isolated Scenedesmaceae strains: a. *T. obliquus*; b. *D. abundans*; c. *T. incrassatulus*; d. *S. quadricauda* C2; e. *D. bicaudatus*; f. *S. quadricauda* C3; g. *D. costato-granulatus*

### 3.2. Biological characteristics of isolated Scenedesmaceae strains

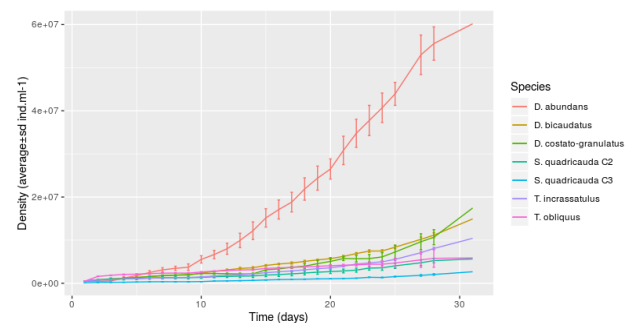


**Figure 2.** Cell density of isolated microalgae strains

In the seven isolated microalgae strains, *D. abundans* showed an outstanding high density ( $60 \times 10^6 \text{ cells/mL}$ ) in comparison with the others, followed by *D. costato-granulatus* ( $17 \times 10^6 \text{ cells/mL}$ ), *D. bicaudatus* ( $15 \times$

$10^6 \text{ cells/mL}$ ), and *T. incrassatulus* ( $10 \times 10^6 \text{ cells/mL}$ ). The three strains *S. quadricauda* C2, *S. quadricauda* C3, and *T. obliquus* had the lowest cell density, which ranged from  $2.6 \times 10^6 - 5.8 \times 10^6 \text{ cells/mL}$  (Figure 2).

Furthermore, the difference in growth characteristics in 31 days among isolated microalgae strains was also recognized. Most of the strains developed very slowly in the first 10 days to get familiar with the culture medium condition (Figure 3). Meanwhile, *D. costato-granulatus* and *S. quadricauda* C2 seemly needed a longer time for their adaptation, which was 15 days. The average growth rate in this period was very low and ranged from  $0.027 - 0.062 \text{ d}^{-1}$ . After that, all of the strains began to grow rapidly that made the cell density at this phase reach very high numbers. *D. abundans* and *S. quadricauda* C2 showed a similar trend in development with the growth rates increased sharply until the 27<sup>th</sup> day and started to show signs of slowing down its growth from the 28<sup>th</sup> day. The growth trend of *D. abundans* in this study was rather different from that in an earlier report of El-Sheekh et al. [8], where *D. abundans* changed to the stationary phase on the 16<sup>th</sup> day. This difference might be due to the differences in strain, culture medium, and isolated condition. The growth curve of the four strains *D. costato-granulatus*, *D. bicaudatus*, *S. quadricauda* C3, and *T. incrassatulus* was different from the other strains with a sharp increase in cell density observing from the 25<sup>th</sup> day and a continuously increasing trend after the 31<sup>st</sup> day. These results once again were not consistent with previous studies such as the result of Tuan [19], where *S. quadricauda* reached the stationary phase on the 16<sup>th</sup> day, or the result of Pena-Castro et al. [20], where *T. incrassatulus* entered the stationary phase on the 14<sup>th</sup> day. Among the isolated strains, only *T. obliquus* entered the stationary phase from the 29<sup>th</sup> day after reaching the highest density ( $5.78 \times 10^6 \text{ cells/mL}$ ) on the 28<sup>th</sup> day, which was a little later than the result of a previous study, where stated that the microalgae started to enter the equilibrium phase on the 21<sup>st</sup> day and showed signs of entering a recession phase on the 30<sup>th</sup> day [6].



**Figure 3.** Growth curves of isolated microalgae strains cultivated in BBM medium (25 °C, 2500 lux, 16:8 day/night lighting time)

In general, in the culture medium condition, *D. abundans* was the strain exhibited the highest growth rate and the shortest generation time among the isolated microalgae strains ( $\mu = 0.160 \text{ d}^{-1}$ ,  $g = 4.34 \text{ days}$ ), followed by *D. costato-granulatus* ( $\mu = 0.118 \text{ d}^{-1}$ ,  $g = 5.58 \text{ days}$ ), *D. bicaudatus* ( $\mu = 0.113 \text{ d}^{-1}$ ,  $g = 6.12 \text{ days}$ ), *S. quadricauda* C3 ( $\mu = 0.109 \text{ d}^{-1}$ ,  $g = 6.33 \text{ days}$ ), and *T. incrassatulus* ( $\mu = 0.101 \text{ d}^{-1}$ ,  $g = 6.58 \text{ days}$ ) (Figure 4).

*S. quadricauda* C2 and *T. obliquus* were the two strains having the lowest average growth rate, which were  $0.081 \text{ d}^{-1}$  ( $g = 8.58$  days) and  $0.082 \text{ d}^{-1}$  ( $g = 8.44$  days), respectively (Figure 4).

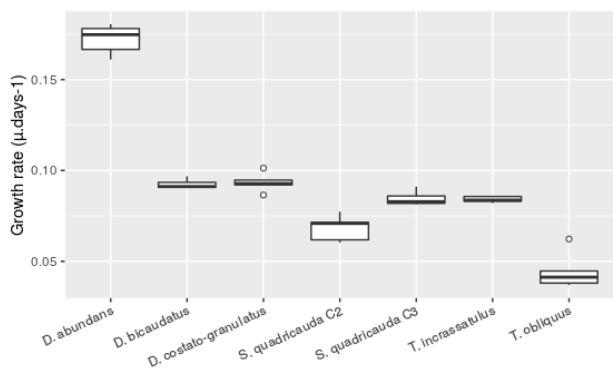


Figure 4. Typical growth rate of isolated microalgae strains

### 3.3. Potential applications of isolated *Scenedesmus* strains

Chlorophyll-a, chlorophyll-b, and carotenoid concentrations of microalgae strains were determined on the 28<sup>th</sup> day of the experiment. Among seven isolated strains, *D. abundans* and *D. costato-granulatus* were the two species having the highest level of both chlorophyll (6.28 and 6.54  $\mu\text{g/mL}$  for chlorophyll-a, and 3.57 and 3.00  $\mu\text{g/mL}$  for chlorophyll-b, respectively) and carotenoid (1.45  $\mu\text{g/mL}$  and 1.49  $\mu\text{g/mL}$ ) (Figure 5). It is worth noting that they were also strains with the highest growth rate and density. Interestingly, although the growth rate and density of *D. costato-granulatus* ( $\mu = 0.111 \text{ d}^{-1}$ ;  $17.44 \times 10^6$  cells/mL) was much lower than those of *D. abundans* ( $\mu = 0.16 \text{ d}^{-1}$ ;  $60.17 \times 10^6$  cells/mL), its content of chlorophyll and carotenoid were rather similar and even a little higher. The high growth rate in combination with the high content of chlorophyll and carotenoid of *D. abundans* and *D. costato-granulatus* demonstrated that they are potential strains for the extraction of these two pigment groups.

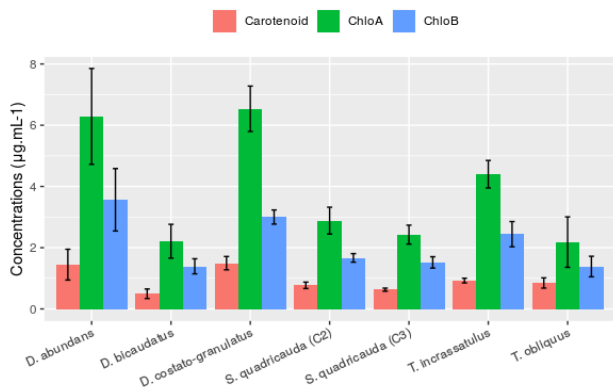


Figure 5. Concentration of chlorophyll-a, chlorophyll-b, and carotenoid in isolated microalgae strains

In terms of potential for biofuel production, *D. costato-granulatus* showed a superiority over the other isolated strains with the lipid content accounting for 74.1% of dry cell weight (dcw) (Figure 6). This was a very high lipid content compared to that of other recorded

Scenedesmaceae strains such as *S. costatus* (10.19% dcw) [21], *S. deserticola* (18.05% dcw) [22], *Scenedesmus* sp. (19.6% dcw) [23], *Coelastrella* sp. (30.8% dcw) [24], *Desmodesmus* sp. (36.7% dcw) [24], as well as that of reported microalgae species with high lipid productivity such as *Chlorella sorokiniana* (34.56% dcw) [21], *C. vulgaris* (34.18% dcw) [24], *Neochlorisoleobundans* (35% - 54% dcw) [23]. Furthermore, it should be noted that this lipid level of *D. costato-granulatus* was obtained in a normal condition of culture medium, and it might increase more if this strain is cultivated in an optimal medium for lipid production, making it become a leading candidate for biodiesel production.

The remaining strains had relatively high lipid contents (28.9% - 43.4% dcw), except *S. quadricauda* C2 (16.6%), but unfortunately, their growth rates were not high ( $0.081 - 0.113 \text{ d}^{-1}$ ). Prominent among these strains was *D. abundans*, which could be considered another potential candidate for lipid production because although its lipid content (32.4% dcw) was not too high in comparison to that of strains in this study, it was higher than that of many microalgae strains in other works [21], [23], and especially, *D. abundans* exhibited an outstanding growth rate ( $\mu = 0.16 \text{ d}^{-1}$ ) among the isolated strains.

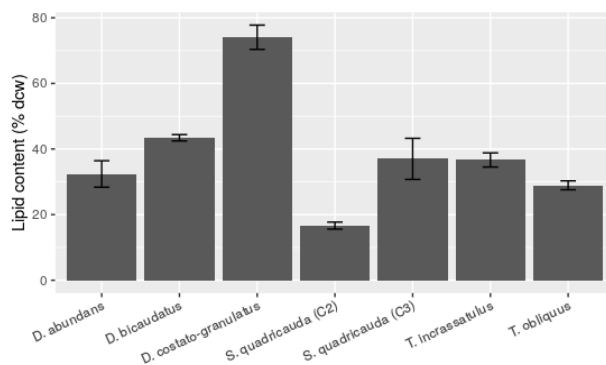


Figure 6. Lipid content in isolated microalgae strains

## 4. Conclusion

In this study, seven microalgae strains belonging to 6 species, 3 genera of the family Scenedesmaceae, division Chlorophyta were isolated and cultivated. In the culture medium condition, these strains showed a growth rate from  $0.081 - 0.160 \text{ d}^{-1}$ . The cell density was rather high but varied among strains, which ranged from  $2.6 \times 10^6 - 60 \times 10^6$  cells/mL. *D. abundans* and *D. costato-granulatus* had the highest growth rate ( $0.160$  and  $0.118 \text{ d}^{-1}$ ) and density ( $60 \times 10^6$  and  $17 \times 10^6$  cells/mL) together with the highest pigment contents (chlorophyll-a: 6.28 and 6.54  $\mu\text{g/mL}$ , chlorophyll-b: 3.57 and 3.00  $\mu\text{g/mL}$ , and carotenoid: 1.45 and 1.49  $\mu\text{g/mL}$ , respectively) and an outstanding lipid (32.4% and 74.1% dcw). Therefore, these two strains could be considered as the most potential strains for chlorophyll and carotenoid extraction and lipid production.

**Acknowledgments:** We would like to thank the Faculty of Biology and Environmental Science, the Faculty of Chemistry, University of Science and Education - The

University of Danang for providing research facilities, and the University of Science and Education, Danang for research funding.

## REFERENCES

- [1] Custódio, L., Soares, F., Pereira, H., Barreira, L., Vizetto-Duarte, C., Rodrigues, M. J., Rauter, A., P., Alberício, F., Varela, J., "Fatty acid composition and biological activities of *Isochrysis galbana* T-ISO, *Tetraselmis* sp. and *Scenedesmus* sp.: possible application in the pharmaceutical and functional food industries", *Journal of Applied Phycology*, 26(1), Springer Link, 2014, 151 - 161.
- [2] Ishan, A., G., Matias-Peralta, H., M., Basri, H., "Bioactive compounds from green microalga - *Scenedesmus* and its potential applications: A brief review", *Pertanika Journal of Tropical Agricultural Science*, 39(1), 2016, 1 - 15.
- [3] Das, M., & Keshri, J., P., "Scenedesmus Meyen & related genera in foot hills of Eastern Himalaya, India", *Phykos*, 45(1), Phycological Society, India, 2015, 75 - 84.
- [4] Shi, J., Podola, B., & Melkonian, "Removal of nitrogen and phosphorus from wastewater using microalgae immobilized on twin layers: an experimental study", *Journal of Applied Phycology*, 19(5), 2007, 417 - 423.
- [5] Chong, A., M., Y., Wong, Y., S., & Tam, N., F., Y., "Performance of different microalgal species in removing nickel and zinc from industrial wastewater", *Chemosphere*, 41(1-2), 2000, 251 - 257.
- [6] Mandal, S. & Mallick, N., "Microalga *Scenedesmus obliquus* as a potential source for biodiesel production", *Appl Microbiol Biotechnol*, 84, Springer-Verlag, 2009, 281-291.
- [7] Piligaev, A., V., Sorokina, K., N., Bryanskaya, A., V., Peltek, S., E., Kolchanov, N., A., & Parmon, V., N., "Isolation of prospective microalgal strains with high saturated fatty acid content for biofuel production", *Algal Research*, 12, Elsevier, 2015, 368 - 376.
- [8] El-Sheekh, M., Abomohra, A., E., Eladel, H., Battah, M., & Mohammed, S., "Screening of different species of *Scenedesmus* isolated from Egyptian freshwater habitats for biodiesel production", *Renewable Energy*, 129, 2018, 114 - 120.
- [9] Tien, D., D. & Hanh, V., *Vietnam freshwater microalgae - Classification of green algae (Chlorococcales)*, Publishing House Academy of Agriculture, Hanoi, 1997.
- [10] San, N., D., "Diversity of green algae (Chlorophyta) in Lake Xuan Duong, Dien Chau district", *Nghe An Journal of Science and Technology*, 6, 2015, 1 - 5.
- [11] Ha, L., T., T., & Trang, B., T., Q., "Composition of green algae (Chlorophyta) in Lake Tau Voi, Ky Anh, Ha Tinh", *Proceedings of the 7th National conference on Ecology and Biological resources*, Vietnam Academy of Science and Technology, 2017, 657 - 663.
- [12] Hanh, H., S., Hanh, V., Son, L., V., "The species composition of green algae (order Chlorococcales) in some estuaries of the Tien and Hau rivers", *Proceedings of the 4th National conference on Ecology and Biological resources*, Vietnam Academy of Science and Technology, 2011, 109 - 114.
- [13] Andersen, R., A., *Algal Culturing Techniques*, Elsevier, 2005.
- [14] Bellinger, E., G. & Sigeo, D., C., *Freshwater Algae: Identification, enumeration and use as bioindicators*, 2nd edition, Wiley Blackwell, 2015.
- [15] Hakalin, N., L., S., Paz, A., P., Aranda, D., A., G., & Moraes, L., M., P., "Enhancement of cell growth and lipid content of a freshwater microalga *Scenedesmus* sp. by optimizing nitrogen, phosphorus and vitamin concentrations for biodiesel production", *Natural Science*, 6(12), 2014, 1044-1054.
- [16] Dung, N., L., Quyen, N., D., Ty, P., V., *Microbiology*, Vietnam education publishing house, 2001.
- [17] Nayek, S., Haque, C., I., Nishika, J., & Suprakash, R., "Spectrophotometric analysis of chlorophylls and carotenoids from commonly grown fern species by using various extracting solvents", *Research Journal of Chemical Sciences*, 4(9), International Science Congress Association, 2014, 63 - 69.
- [18] Bligh, E., G. & Dyer, W., J., "A rapid method of total lipid extraction and purification", *Can. J. Biochem. Physiol.*, 37(8), The National Research Council of Canada, 1959, 911 - 917.
- [19] Tuan, D., V., 2014, *Isolation and investigation of resistance to some bacterial and fungal pathogens of Scenedesmus quadricauda*, Hanoi University of Agriculture, 2014.
- [20] Peña-Castro, J., M., Martínez-Jerónimo, F., Esparza-García, F., & Cañizares-Villanueva, R., O., "Phenotypic plasticity in *Scenedesmus incrassatulus* (Chlorophyceae) in response to heavy metals stress", *Chemosphere*, 57(11), 2004, 1629 - 1636.
- [21] Thao, T., Y., Linh, D. T., N., Sy, V. C., Chau, T., N., M., Khai, N., N., & Thao, P., D., P., "Isolation of lipid producing microalgae from fresh, brackish and marine water sources", *Journal of Science*, 3(81), Ho Chi Minh city University of Education, 2016, 88 - 99.
- [22] Thanh, P., D., Hau, T., T., & Toan, D., T., "The effect of nitrogen or phosphorus deficiency on the lipid accumulation in micro-algae *Scenedesmus Desertiicola*", *Journal of Science*, 16(4), An Giang University, 2017, 40 - 47.
- [23] Prabhakaran, M., Sivasankar, V., Omine, K., & Vasanthi, M., "Microalgae biofuels: a green renewable resource to fuel the future", *Green Technologies and Environmental Sustainability*, Springer International Publishing, 2017, 105 - 129.
- [24] Ferro, L., Gentili, F., G., & Funk, C., "Isolation and characterization of microalgal strains for biomass production and wastewater reclamation in Northern Sweden", *Algal Research*, 32, Elsevier B.V., 2018, 44-53.

(The Board of Editors received the paper on 18/12/2019, its review was completed on 06/02/2020)